

REVIEW ARTICLE

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# Mechanistic Insights into the Efficacy of Sodium Bicarbonate Supplementation to Improve Athletic Performance

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## Abstract

A large proportion of empirical research and reviews investigating the ergogenic potential of sodium bicarbonate ( $\text{NaHCO}_3$ ) supplementation have focused predominately on performance outcomes and only speculate about underlying mechanisms responsible for any benefit. The aim of this review was to critically evaluate the influence of  $\text{NaHCO}_3$  supplementation on mechanisms associated with skeletal muscle fatigue as it translates directly to exercise performance. Mechanistic links between skeletal muscle fatigue, proton accumulation (or metabolic acidosis) and  $\text{NaHCO}_3$  supplementation have been identified to provide a more targeted, evidence-based approach to direct future research, as well as provide practitioners with a contemporary perspective on the potential applications and limitations of this supplement. The mechanisms identified have been broadly categorised under the sections 'Whole-body Metabolism', 'Muscle Physiology' and 'Motor Pathways', and when possible, the performance outcomes of these studies contextualized within an integrative framework of whole-body exercise where other factors such as task demand (e.g. large vs. small muscle groups), cardio-pulmonary and neural control mechanisms may outweigh any localised influence of  $\text{NaHCO}_3$ . Finally, the 'Performance Applications' section provides further interpretation for the practitioner founded on the mechanistic evidence provided in this review and other relevant, applied  $\text{NaHCO}_3$  performance-related studies.

## Key Points

- $\text{NaHCO}_3$  has wide-ranging effects on mechanisms related to whole-body metabolism, specifically the [PCr]/[Pi]-power relationship, glycolytic intermediates and the intra- and extracellular distribution of metabolites and other strong ions.
- $\text{NaHCO}_3$  may affect muscle physiology and motor pathways associated with early rate of force development and/or the metabolic properties associated with contractile shortening velocity, and this effect appears more pronounced in fast-twitch fibres.
- Appropriately identifying whether  $\text{NaHCO}_3$  ingestion may influence the underlying mechanisms associated with whole-body metabolism, muscle physiology and/or motor pathways within the context of a particular athletic

endeavour or training stimulus is an important initial step when considering the use of this supplement.

## Review

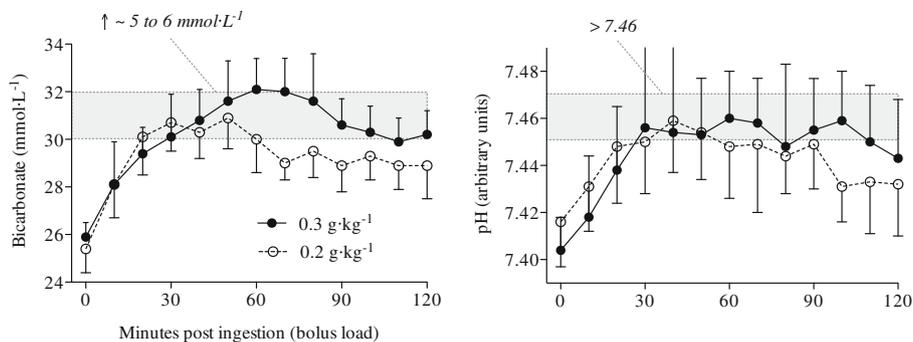
### Background

For over 40 years, researchers have explored the efficacy of inducing alkalosis to enhance athletic performance. Although many buffers have been studied (e.g. sodium citrate, sodium phosphate, sodium lactate), evidence supports sodium bicarbonate ( $\text{NaHCO}_3$ ) as the most consistently effective agent for improving exercise performance. Meta-analyses have reported that supplementation can result in an approximately 2 to 3 % improvement in a variety of performance measurements (e.g. power, speed, work capacity, time to failure) during both single and repeated bouts of high-intensity exercise [1, 2]. Oral administration of  $\text{NaHCO}_3$  generally increases blood buffering capacity (Fig. 1) [3] and is believed to attenuate the increase in intramuscular acidity

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**Fig. 1** Resting changes in blood bicarbonate [ $\text{HCO}_3^-$ ] and pH after ingesting 0.2 and 0.3  $\text{g}\cdot\text{kg}^{-1}$  of sodium bicarbonate ( $\text{NaHCO}_3$ ) (reproduced from Siegler et al. [3] with permission)

synonymous with high-intensity exercise and skeletal muscle fatigue [1, 4], although the physiologic mechanisms directly responsible for performance augmentation in humans are unclear. Indeed, a large proportion of empirical research and reviews investigating the ergogenic potential of  $\text{NaHCO}_3$  focus predominately on performance outcomes (e.g. time to task failure, cumulative work accomplished, time trial performance) [1, 4, 5], and only speculate about underlying mechanisms responsible for attenuating skeletal muscle fatigue.

Mechanistic links between  $\text{NaHCO}_3$  supplementation, skeletal muscle fatigue and athletic performance are often underpinned by the belief that metabolic acidosis, or a decline in muscle pH, contributes to skeletal muscle fatigue through inhibition of various metabolic processes and/or rates of contractile cycling [6–9]. However, it has also been argued that acidosis has little influence on maximal force production [10], and in some instances may even facilitate maximal force and velocity of shortening [11]. The debate as to whether or not metabolic acidosis, or indeed the intra- and extracellular distribution of other relevant ions (e.g.  $\text{Ca}^{2+}$ ,  $\text{K}^+$ ,  $\text{Cl}^-$ ), has an inhibiting or facilitating effect on contractile function continues today and has been summarised in recent reviews [12–14]. The primary aim of this review was to fundamentally examine the influence of  $\text{NaHCO}_3$  supplementation on metabolic acidosis and mechanisms associated with skeletal muscle fatigue and athletic performance. Secondly, the mechanistic evidence presented in this review has been contextualised in the ‘Performance Applications’ section to provide contemporary, evidence-based recommendations for practitioners and athletes.

After the ‘Historical Overview’ section, studies have been broadly categorised under sub-headings, and when possible, the performance outcomes of these studies contextualized within an integrative framework of whole-body exercise where other factors such as task demand (e.g. large vs. small muscle groups), cardio-pulmonary and

neural control mechanisms may outweigh any localised influence of  $\text{NaHCO}_3$ . In many of these studies,  $\text{NaHCO}_3$  is also used in conjunction with an acid (e.g. ammonium chloride ( $\text{NH}_4\text{Cl}$ )) to determine the wider contribution of acid-base perturbation on skeletal muscle fatigue; however, emphasis has been placed on reviewing only the alkalosis condition in most instances. This review does not address all performance-based applications of  $\text{NaHCO}_3$  supplementation in the literature, such as team sport performance [15–17], skill sports [18–20], hydration [21, 22] or the efficacy of  $\text{NaHCO}_3$  under hypoxic conditions [23–25].

#### Historical Overview

Associations between  $\text{NaHCO}_3$ , skeletal muscle fatigue and exercise performance can be traced back to what is believed to be the first paper published on the topic in 1931 from the Harvard Fatigue Laboratory [26]. This study was one of many in this era more broadly focused on respiratory and metabolic buffering mechanisms associated with what was termed ‘lactic acidosis’ [26]. Indeed, there was considerable interest in lactic acidosis and fatigue during muscle contractions and exercise as early as the 1900s and continuing through to the 1970s [27]. However, beyond a few subsequent investigations [28–31], research on the ergogenic potential of  $\text{NaHCO}_3$  did not become prominent until the mid-1970s.

The first series of exercise performance studies to be formally presented from one laboratory were from Jones and colleagues starting in 1977 [32–35]. Collectively, these were also the first studies on humans to incorporate both an alkali ( $\text{NaHCO}_3$ ) and an acid ( $\text{NH}_4\text{Cl}$ ) to explore the impact of altered acid-base status on markers of cardiovascular, respiratory and metabolic function in conjunction with exercise performance. In the first two studies, cycling at 95 % of  $\text{VO}_{2\text{max}}$  to volitional exhaustion after  $\text{NaHCO}_3$  ingestion elicited between a 30 and 40 % performance improvement, respectively [32, 33], whereas a following study requiring a 30-s maximal

effort at 100 rpm only resulted in a small but non-significant improvement in maximal power [34]. In the fourth study, incorporating an incremental cycling protocol ( $\sim 16 \text{ W min}^{-1}$  at 60 rpm to volitional exhaustion), only the  $\text{NH}_4\text{Cl}$  condition induced a performance decrement whereas there was no difference between the  $\text{NaHCO}_3$  and control conditions [35]. The authors accounted for the performance discrepancies by relating exercise task and duration to a decline in muscle pH and the concurrent effect on the rate of glycolysis. Furthermore, they suggested that the beneficial effects of enhanced  $\text{H}^+$  removal would be more advantageous during the prolonged cycling tasks (e.g.  $>4$  min) than during the shorter, 30-s efforts [34]. The discrepancy in performance outcomes in these studies is also the first of many examples where different exercise task demands and  $\text{NaHCO}_3$  dosing protocols may have impacted upon performance outcomes.

From the mid-1980s, performance-related  $\text{NaHCO}_3$  studies began to emerge prominently in the literature. The increased interest may have been influenced by earlier findings [32, 33], but was probably also related to the ease of oral  $\text{NaHCO}_3$  administration and the short time course required to acutely induce metabolic alkalosis that provided the opportunity to conduct a wide variety of performance-related studies within a relatively short period of time [36–40]. Collectively, the studies during this period that reported an ergogenic benefit of  $\text{NaHCO}_3$  ingestion generally adhered to a maximal or supramaximal (e.g. 125 %  $\text{VO}_2\text{max}$ ) exercise task lasting between 1 and 7 min (Table 1) [41, 42]. Many of the performance-based studies in this era also proposed a similar study rationale; one based on previous equivocal performance outcomes and/or on the attenuation of the decline in muscle pH proposed to occur during high rates of glycolytic flux after  $\text{NaHCO}_3$  ingestion [39, 40, 43–45]. Indeed, these rationales persist in many published papers today [46–49], further illustrating that

beyond the effect on glycolysis, the other potential physiologic mechanisms responsible for performance augmentation after  $\text{NaHCO}_3$  supplementation have received limited attention in the applied research domain.

### Whole-Body Metabolism

#### *Anaerobic/Aerobic Capacity*

From an energetic perspective, any physiological effects of  $\text{NaHCO}_3$  supplementation that translate to an increase in performance should be observed in the rates and amounts of energy provision and/or the efficiency of energy transduction to power and work. However, very little research has investigated, or indeed attempted to quantify, anaerobic and aerobic energy yields, accumulated oxygen deficits or energetic efficiency during conditions of metabolic alkalosis. Beyond a handful of studies that have modelled whole-body  $\text{VO}_2$  kinetics (e.g.  $\text{VO}_2$  fast and slow components [50–52]), most of the research on  $\text{NaHCO}_3$  and metabolism has been conducted on single muscle or isolated muscle groups and their metabolic response to various task demands.

From studies conducted in the 1970s [32, 53–55], a causal link between increased intramuscular lactate ( $\text{La}^-$ ) concentrations and  $\text{La}^-$  efflux, reduced proton ( $\text{H}^+$ ) accumulation and improved muscular performance had been posited to explain the performance improvements observed following  $\text{NaHCO}_3$  supplementation [6]. To further clarify the influence of  $\text{NaHCO}_3$  on intramuscular [ $\text{La}^-$ ] and  $\text{La}^-$  efflux during intense exercise, Spriet and colleagues minimised the limitations associated with in vivo contracting muscle measurement (e.g. varying work rates and metabolite accumulation) by applying continuous supramaximal stimulation (5 V at 0.5 Hz) for 20 min to a perfused gastrocnemius-plantaris-soleus (GPS) muscle group [56].  $\text{NaHCO}_3$  increased the rate of  $\text{La}^-$  efflux from the muscle group, but did not attenuate the decline in muscle tension as compared to control conditions [56]. Methodological limitations inherent to the perfusion and

**Table 1** A summary of studies conducted in the mid 1980's that reported an ergogenic effect of  $0.3 \text{ g kg}^{-1}$  sodium bicarbonate ( $\text{NaHCO}_3$ ) supplementation on performance tasks lasting approximately  $\sim 7$  min

Study	Subject characteristics	Task	Control	$\text{NaHCO}_3$	Improvement (%)
Wilkes et al. '83 [36]	University track athletes	800-m foot race	2:05.8 $\pm$ 2.1 (mins)	2:02.9 $\pm$ 1.9 (mins)	$\sim 2$ %
Costill et al. '84 [40]	$\text{VO}_2\text{max}$ : 4.8 $\text{L min}^{-1}$ (3.12–6.33)	5, 1-min cycling bouts (125 % $\text{VO}_2\text{max}$ ; 5th bout to exhaustion)	113.5 $\pm$ 12.4 s	160.8 $\pm$ 19.1 s	$\sim 42$ %
McKenzie et al. '86 [38]	$\text{VO}_2\text{max}$ : 3.83 $\pm$ 0.61 $\text{L min}^{-1}$	6, 60-s cycling bouts (125 % $\text{VO}_2\text{max}$ ; 6th bout to exhaustion)	74.7 $\pm$ 17.6 s	106 $\pm$ 6.9 s	$\sim 30$ %
Gao et al. '86 [37]	Well-trained college swimmers ( $\text{VO}_2\text{max}$ : 4.3 $\pm$ 0.1 $\text{L min}^{-1}$ )	5, 100-yd freestyle swim	4th bout: $\sim 1.65 \text{ m s}^{-1}$ 5th bout: $\sim 1.64 \text{ m s}^{-1}$	4th bout: $\sim 1.62 \text{ m s}^{-1}$ 5th bout: $\sim 1.61 \text{ m s}^{-1}$	$\sim 2$ % $\sim 2$ %
Goldfinch et al. '88 [39]	Male athletes	400-m foot race	58.46 $\pm$ 2.49 s	56.94 $\pm$ 2.25 s	$\sim 3$ %

Subject characteristics are represented to the level of detail provided in the original published studies. All values in the (%) improvement column were significantly different from control ( $p < 0.05$ )

stimulation protocol only allowed the authors to speculate as to whether or not the increased  $\text{La}^-$  efflux was a result of increased rates of glycogenolysis and/or glycolysis, or due to an increased rate of  $\text{La}^-$  release from the muscle.

Nearly 15 years later, Hollidge-Horvat and colleagues used human muscle tissue samples obtained at different steady-state exercise intensities (i.e. 30, 60 and 75 %  $\text{VO}_2\text{max}$ ) to investigate the mechanisms responsible for the increased  $\text{La}^-$  efflux by then commonly observed after  $\text{NaHCO}_3$  ingestion [57]. These authors eloquently detailed a complex, intensity-dependent response in key metabolic enzymes and regulatory steps after  $\text{NaHCO}_3$  ingestion as compared to control conditions. Briefly, the authors observed a similar increase in  $\text{La}^-$  efflux to that of previous studies after  $\text{NaHCO}_3$  ingestion, but only at 75 %  $\text{VO}_2\text{max}$  [57]. The authors speculated that the increased  $[\text{La}^-]$  observed in circulation may have been due to a decrease in  $\text{La}^-$  uptake by inactive tissue [58] or an altered  $\text{H}^+$  gradient on the transporters (later confirmed by others as an increase in monocarboxylate transporter (MCT) activity [59]). The authors also documented an increased degradation of muscle glycogen in the  $\text{NaHCO}_3$  compared to the control trials [57]. As performance was not assessed in this study, whether or not this increased reliance on muscle glycogen stores in the  $\text{NaHCO}_3$  trial would have influenced performance at the higher intensities remains unknown.

As the capacity to study exercising muscle metabolism in vivo improved beyond the limitations of the biopsy, others have subsequently expanded upon these findings to include exercise performance measures. Using phosphorus-31 magnetic resonance spectroscopy ( $^{31}\text{P}$ -MRS) and a constant-rate progressive wrist-flexion exercise (1-s contraction/1-s relaxation) to volitional exhaustion, Raymer and colleagues reported an approximate 12 % increase in time to failure after  $\text{NaHCO}_3$  supplementation and attributed the performance effect to a delayed onset of intracellular acidosis (evident at ~60 % peak power) in the alkalotic condition [60]. Furthermore, concomitant to the delayed intracellular acidosis was a delay in the slow-to-rapid increase in the  $[\text{PCr}]/[\text{Pi}]$  to power output relationship (repeatedly observed in subsequent studies by this same group [61, 62]). As metabolic perturbation caused by acidosis had been associated with the inhibition of glycolysis and glycogenolysis [63], increased phosphocreatine (PCr) degradation and inorganic phosphate ( $\text{P}_i$ ) accumulation ( $[\text{PCr}]/[\text{P}_i]$ ) within the myocyte [64], the authors speculated that the delayed accumulation of intracellular  $\text{H}^+$  induced by  $\text{NaHCO}_3$  was the primary cause of the performance effect.

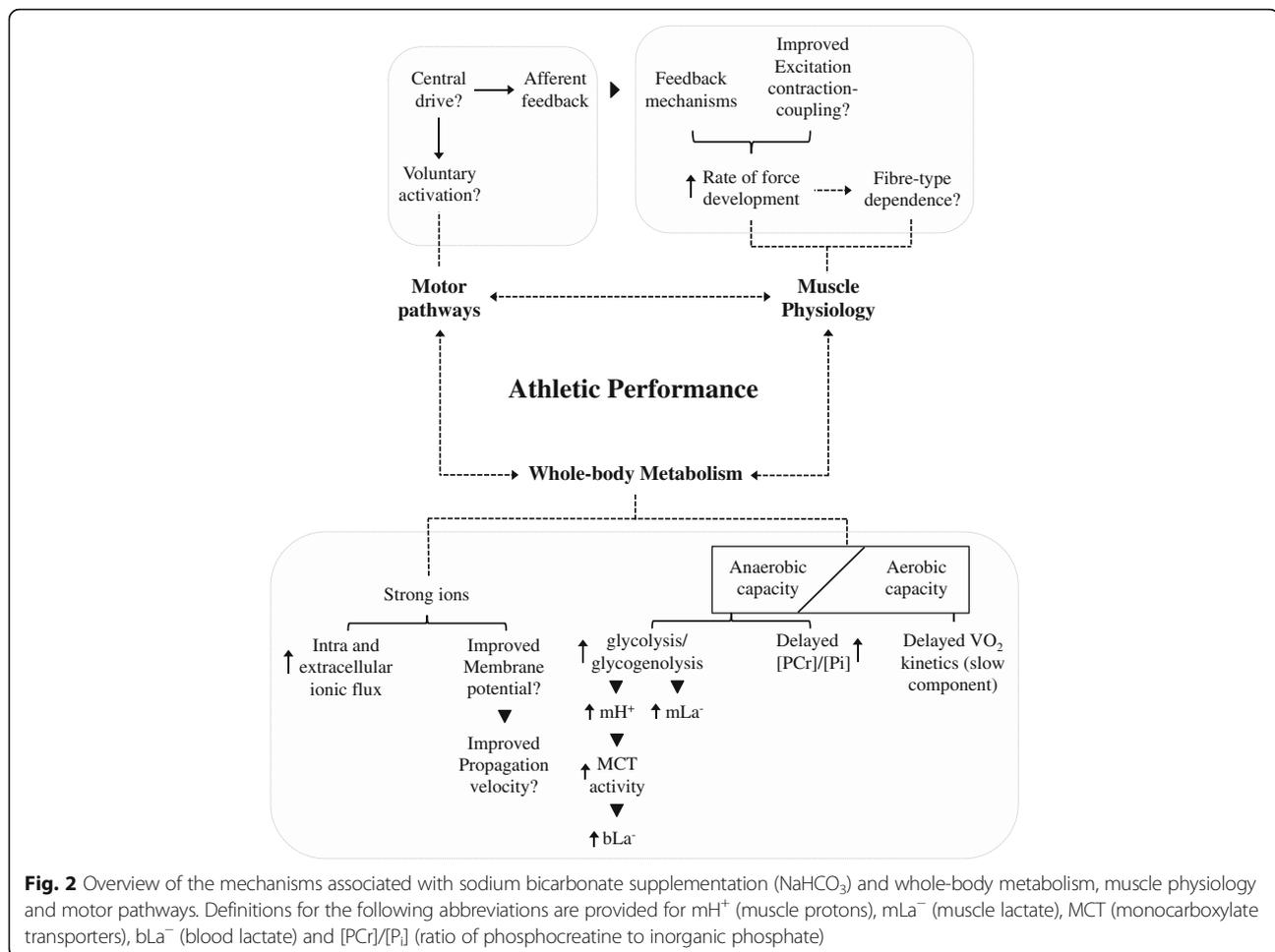
To briefly summarise, much of the research in this area has been centred around muscle metabolism, from

the earlier observations on intramuscular  $[\text{La}^-]$  and  $\text{La}^-$  efflux to the more recent focus on glycolytic intermediates and high-energy phosphate kinetics (e.g. PCr).  $\text{NaHCO}_3$  appears to effect the  $[\text{PCr}]/[\text{Pi}]$ -power relationship, glycolytic intermediates and the intra- and extracellular distribution of metabolites and other strong ions (Fig. 2) [56, 57, 60, 65, 66]. Not surprisingly, this effect is more pronounced during periods of prolonged, intense contractile activity where a larger proportion of energy provision is derived from classically defined *anaerobic pathways*. Contextually, this effect may also partially explain the ergogenic findings of many performance-based studies on athletes that have imposed a high demand on these pathways either with single or repeated bouts of intense exercise [39, 67–70].

### Strong Ions

Inducing a state of alkalosis also affects the intra- and extracellular balance of other strong ions such as  $\text{Na}^+$ ,  $\text{K}^+$  and  $\text{Cl}^-$  [60], all of which contribute to the maintenance of skeletal muscle contractile function [13]. In general, and irrespective of acid-base balance, fatiguing exercise induces ion concentration changes between the intracellular, interstitial and extracellular compartments. Although ultimately dependent upon the level of contractile activity, as well as diffusion and perfusion limitations, muscle  $\text{K}^+$  will tend to decrease whereas muscle  $\text{Na}^+$ ,  $\text{Cl}^-$  and  $\text{Ca}^{2+}$  will tend to increase with intense exercise ( $\text{H}^+$  will increase in both intra- and extracellular compartments) [13]. Collectively, changes in these ionic concentrations have been demonstrated to decrease resting membrane potential and impair sarcolemmal excitability [13]. The effect of  $\text{NaHCO}_3$  specifically on the distribution of these ions between compartments, in particular the  $\text{NaHCO}_3^-$ -induced reduction in  $\text{K}^+$  efflux from the muscle, has been the focus of other studies investigating the effect of alkalosis on the interstitial and plasma concentrations of strong ions and skeletal muscle fatigue [65, 66, 71].

In terms of exercise performance, however, only Sosteric and colleagues have demonstrated an improvement after  $\text{NaHCO}_3$  supplementation [65]. By incorporating a small muscle mass, and therefore inducing a larger reduction in muscle  $\text{K}^+$  than commonly observed during whole-body exercise [72], the authors hypothesised that  $\text{NaHCO}_3$  would attenuate the augmented  $\text{K}^+$  efflux from the small muscle mass and therefore better preserve muscle function [65]. Subsequently, they observed an increase in total exercise time of greater than 2 min (~25 % improvement) compared to a placebo during a constant rate, progressive concentric forearm flexion exercise task to volitional exhaustion [65].  $\text{NaHCO}_3$  also improved the regulation of circulating  $\text{K}^+$  and handling of  $\text{Cl}^-$  (but not  $\text{Na}^+$ ), which the authors proposed to



have better preserved membrane excitability and therefore contributed to the performance improvement [65].

$\text{NaHCO}_3$  appears to delay the rise in muscle interstitial  $\text{K}^+$  that occurs during intense contractile activity and, via this effect, may attenuate the decline in muscle membrane excitability and the extent of  $\text{K}^+$ -induced inactivation of myocytes during such activity [11, 66]. As previously stated, this effect may have contributed to not only the performance improvement observed by Sostaric and colleagues but also the earlier findings of Raymer as both used a similar exercise performance task [60, 65]. However, these studies also illustrate the relevance of muscle mass and task demand when determining the efficacy of  $\text{NaHCO}_3$  as an ergogenic supplement. Issues such as agonist/antagonist muscle actions, synergistic contraction of larger muscle groups and recruitment order (e.g. Hennemen's size principle) [73] during whole-body exercise may mitigate the performance effects observed in smaller, isolated muscles. As an example, the trend of  $\text{NaHCO}_3$  improving incremental exercise to volitional exhaustion in smaller muscle groups [60, 65] is less consistent when large muscle

groups (e.g. cycling) or whole-body exercise is incorporated within a similar task [32, 33, 35, 74, 75] (Table 2).

## Muscle Physiology

### Influence of Fibre-Type

Although earlier work had illustrated a relationship between muscle pH and skeletal muscle force and/or muscle shortening velocity [76–78], the first to show an independent effect of acidosis on fast- and slow-twitch fibres appears to be Metzger and Moss in 1987 [79]. Using single fibres of rat soleus (SOL) and vastus lateralis (VL) muscles, previously characterised as predominately type I (SOL) or IIa and b (VL), respectively [80, 81], maximal isometric tension and maximum shortening velocity was assessed as a function of pH. Although initial declines in pH induced a concomitant reduction in tension and shortening velocity in both SOL and VL, as pH decreased below 6.2, the relative loss in tension was greater in VL [79]. The first to study the effect of  $\text{NaHCO}_3$  on fibre-specific force production was Lindinger and colleagues in 1990 [82]. These authors examined the effects of  $\text{NaHCO}_3$  on SOL and white gastrocnemius (WG) muscle tension at

**Table 2** A summary of the results of studies investigating the influence of 0.3 g kg<sup>-1</sup> sodium bicarbonate (NaHCO<sub>3</sub>) supplementation on whole-body and isolated muscle incremental exercise to volitional exhaustion

Study	Whole-body or isolated muscle	Incremental task	Performance effect
Raymer et al. '04 [60]	Isolated (forearm)	1:1 contraction-relaxation cycle; initial resistance 1.0 kg then increased 0.22 kg min <sup>-1</sup>	~12 % improvement
Sostaric et al. '05 [65]	Isolated (concentric finger flexion)	30 rep min <sup>-1</sup> ; initial resistance 2.6 ± 0.22 W then increased 0.17 W min <sup>-1</sup>	~25 % improvement
Poulus et al. '74 [74]	Whole-body (cycling)	Fixed cadence with 10 W min <sup>-1</sup> increased until volitional exhaustion)	No effect
Jones et al. '77 [32]	Whole-body (cycling)	20 min at power output ~33 % VO <sub>2</sub> max, then 20 min at 66 % VO <sub>2</sub> max, then 95 % VO <sub>2</sub> max to volitional exhaustion	~40 % improvement
Sutton et al. '81 [33]	Whole-body (cycling)	20 min at power output ~33 % VO <sub>2</sub> max, then 20 min at 66 % VO <sub>2</sub> max, then 95 % VO <sub>2</sub> max to volitional exhaustion	~30 % improvement
Kowalchuk et al. '84 [35]	Whole-body (cycling)	Increased 16 W min <sup>-1</sup> at 60 rpm	No effect
Housh et al. '91 [75]	Whole-body (cycling)	Continuous (2-min stages) physical work capacity test (PWC <sub>FT</sub> ) starting at 60 W to volitional exhaustion	No effect

All values represented as having an improvement (%) were significantly different from control or placebo ( $p < 0.05$ )

rest and during 5 min of intense stimulation [82]. Although NaHCO<sub>3</sub> differentially altered the ionic composition (specifically K<sup>+</sup>, Cl<sup>-</sup> and Mg<sup>2+</sup>) and the rate of La<sup>-</sup> efflux from the muscle fibres, it had no effect on muscle tension in either muscle [82].

In 2007, Broch-Lips and colleagues stimulated the SOL and extensor digitorum longus (EDL), another muscle characterised as predominately type IIa, under normal or alkalotic conditions. In addition to the isometric force measurement, the authors also estimated rate of force development (RFD) as the maximal numerical slope value of the force development during contraction [71]. Although NaHCO<sub>3</sub> induced increases in pH from 7.4 to 7.6, neither maximum force nor RFD was affected in either muscle [71]. However, more recent evidence using a fatiguing, mechanically controlled stretch-shortening work cycle model has shown that NaHCO<sub>3</sub> administration may improve RFD [83]. Avoiding the limited degree of shortening that occurs during isometric contractions [71], Higgins and colleagues subjected both SOL and EDL mouse muscles to repeated length changes matched to the expected stride frequency of the mice during normal locomotion (10 % strain with a cycle frequency for EDL of 8 Hz and SOL of 5 Hz) and demonstrated an approximate 7 % improvement in force production during shortening in the EDL compared to a 3 % improvement in the SOL after NaHCO<sub>3</sub> incubation.

To date, the fibre-type dependent findings related to contractile properties have not been replicated in the exercising human given the methodological constraints addressed earlier of isolating fibre-type contributions to whole-body locomotion [84], quantifying properties of diffusion and other factors associated with intramuscular blood supply [85, 86]. However, collectively, this body of research suggests that NaHCO<sub>3</sub> may affect the

metabolic properties associated with contractile shortening velocity and that this effect appears more pronounced in fast-twitch fibres (Fig. 2). The effect on rates of contractile shortening but not maximal force may also explain the limited efficacy of NaHCO<sub>3</sub> observed in performance-based studies only measuring maximal voluntary force [87, 88].

#### Chronic Adaptation

Interestingly, although shortening velocity of slow-twitch fibres after acute NaHCO<sub>3</sub> administration does not appear to be improved when compared to fast-twitch fibres, chronic ingestion coupled with high-intensity training may improve the oxidative capacity of these fibres [89–91]. After 8 weeks (three times per week) of chronic NaHCO<sub>3</sub> ingestion coupled with high-intensity (between 140 and 170 % lactate threshold (LT)) intermittent training, Edge and colleagues reported a significant improvement in both LT (~11 %) and performance (~25 %) (time to failure) in moderately trained females when compared to a control group matched for total work (k). The authors speculated that as H<sup>+</sup> accumulation has been shown to impair oxidative capacity and various components of cellular respiration [57], training in an alkalotic state may have attenuated the impairment and allowed for prolonged cellular respiration to occur during training [89].

Continuing this line of investigation, two subsequent studies by the same group used relatively similar NaHCO<sub>3</sub> dosing and high-intensity training protocols on rats to explore fibre-type dependent protein expression associated with H<sup>+</sup> removal during rest and exercise [90, 91]. In the first study, the authors observed a differential effect of alkalosis on MCT-4 expression in the SOL but not EDL when compared to

placebo and control conditions [90]. Additionally, citrate synthase concentration was higher in the alkalotic condition (23 vs. 16 %) after training in the SOL but not EDL. These findings led the authors to conclude that chronic  $\text{NaHCO}_3$  ingestion was influencing protein expression associated with slow but not fast-twitch fibres. The final study in the series appeared to further confirm this observation, with the authors reporting an increase in mitochondrial respiration in the SOL as compared to the EDL after training in an induced alkalotic state [91].

The evidence that chronic use of  $\text{NaHCO}_3$  coupled with appropriate training may lead to aerobic adaptations associated with improved mitochondrial efficiency in slow-twitch fibres is intriguing [91], and future work may clarify whether this is indeed a viable option for improving aerobic efficiency. Additionally, we are unaware of any studies that have investigated the efficacy of introducing chronic  $\text{NaHCO}_3$  supplementation on mechanisms associated with muscle force production or rapid force-generating capacity. Given the preliminary evidence that  $\text{NaHCO}_3$  may exhibit independent effects on fibre-specific mechanisms associated with improved contractile function both acutely and chronically, future study in this area is warranted.

### Motor Pathways

Ultimately, determining the mechanisms by which  $\text{NaHCO}_3$  supplementation can improve athletic performance may require a more integrative understanding of other systems inherently involved during exercise, and more specifically the progression of skeletal muscle fatigue. The motor pathways, or more explicitly the neuromuscular system, are intimately involved in maintaining skeletal muscle function during fatiguing exercise. This complex integrative network is often overlooked in the  $\text{NaHCO}_3$  literature as a possible contributor to whole-body exercise performance. Although partially due to the limited number of studies observing the influence of  $\text{NaHCO}_3$  on the motor pathways, studies in this section have been broadly categorised as they relate to motor unit recruitment and feedback mechanisms associated with metabolite accumulation in the periphery that have been directly linked to central drive.

As addressed in the 'Whole-body Metabolism' section, fluctuations in strong ions (e.g.  $\text{K}^+$ ,  $\text{Na}^+$ ,  $\text{Cl}^-$ ) have also been linked with maintaining resting membrane potential and sarcolemmal excitability [13]. However, few studies have explored whether inducing pre-exercise alkalosis via  $\text{NaHCO}_3$  ingestion influences these mechanisms as they relate to excitation-contraction coupling and ultimately muscle force generating capacity [92–95]. Hunter and colleagues investigated the effects of  $\text{NaHCO}_3$  ingestion on muscle fibre conduction velocity

after fatiguing exercise (i.e. 50 min of cycling at 105 % lactate threshold) [93]. Using a previously developed technique [96], these authors observed an attenuation in the decline in muscle fibre conduction velocity after cycling in an alkalotic state [93]. This improvement, however, did not translate into an improvement in maximal knee extension force.

We have also demonstrated that  $\text{NaHCO}_3$  has no effect on maximal force production [8]. However, we have observed an improvement in the ability to rapidly generate force (RFD), a functional measure related to both rapid motor unit recruitment and contractile shortening velocity [97]. In this study, subjects performed an intermittent (30 s:30 s work-to-rest ratio) cycling protocol at 120 % of peak power until volitional exhaustion, where maximal voluntary contractions (MVC) were performed immediately after each 30-s work cycle. Although RFD and maximal force declined in both  $\text{NaHCO}_3$  and control trials, RFD was better maintained throughout the exercise under alkalotic conditions [97]. The mechanisms responsible for the differential effect on RFD but not maximal force after  $\text{NaHCO}_3$  ingestion requires further investigation, as RFD is influenced by both peripheral (e.g. excitation-contraction coupling) and central (e.g. central drive) factors [98].

Mechanisms related to central drive such as cortical output, spinal reflexes and muscle afferents have also been shown to be influenced by the localised changes in muscle metabolite concentrations (e.g.  $\text{H}^+$ ) associated with exercise [99–101]. The complex interaction between these mechanisms as they contribute to skeletal muscle fatigue has been previously reviewed [99]. Briefly, they are often categorised as supraspinal (e.g. output of descending motor cortical and corticospinal pathways to the motoneurons) and sub-spinal (e.g. muscle spindles, Golgi tendon organs, small diameter group III/IV afferents) mechanisms [99] and their relative contribution to skeletal muscle fatigue associated with the intensity of the exercise task. The degree to which  $\text{H}^+$  accumulation in the periphery affects central mechanisms, particularly during intense exercise, has been related specifically to the central projection of the mechanically and chemically sensitive small diameter group III/IV afferents located within the skeletal muscle [102, 103]. It is believed that the exercise-induced actions of these afferent fibres inhibit the excitability and net output of corticospinal cells in the central nervous system, as well as effect alpha motor neuron excitability [104–107].

Very few investigations have used  $\text{NaHCO}_3$ , or indeed other methods, to manipulate pH and to explore the effects of alkalosis on afferent or other neural pathways contributing to skeletal muscle fatigue [94, 95]. Based on the assumption that  $\text{H}^+$  accumulation leads to a decline in central drive to the muscle, Matsuura and colleagues

measured surface electromyography (sEMG) activity in the vastus lateralis to determine whether motor unit recruitment strategies during repeated high-intensity cycling (i.e. repeated 10-s efforts at a resistive load ( $N$ ) of  $0.075 \text{ BM } 9.81^{-1}$ ) would be influenced by  $\text{NaHCO}_3$  ingestion [94]. Neither root mean square or mean power frequency was influenced by pH, with both  $\text{NaHCO}_3$  and control conditions reflecting similar recruitment profiles throughout the test [94]. A follow-up study by the same group suggested that  $\text{NaHCO}_3$  again had no effect on gross sEMG activity or perceived sense of effort after 2 min of cycling at 105–110 % of a pre-determined maximal workload [95]. However, sEMG signals lack sensitivity to detect changes in the number and discharge rate of active motor units that would indicate increased central motor output [108, 109] and therefore may not be sensitive enough to accurately determine whether or not altering pH may affect descending central drive.

A more recent body of literature suggests a relationship between increasing levels of intramuscular metabolite concentrations analogous with high rates of skeletal muscle contraction, and greater group III/IV afferent firing in humans [110–112], which would ultimately act as a protective mechanism against peripheral fatigue by limiting descending central drive [113]. Using an isometric knee extension model in combination with ischemia to enhance the firing frequency of group III/IV afferents [114–116], we have observed voluntary activation, a gross estimate of central drive, to be better preserved under  $\text{NaHCO}_3$  conditions [117]. The improved activation during alkalosis, however, was not commensurate with changes in net muscle excitation or rapid and maximal force output which declined equally in both  $\text{NaHCO}_3$  and control conditions [117]. We speculated that the divergence might have been a result of the extreme conditions of peripheral fatigue induced by our exercise protocol (2-min MVC of the quadriceps followed by 2 min of ischemia and ending with a 1-min ischemic MVC) [117]. The findings may also reflect the relatively minor role afferent feedback may play in these situations of extreme peripheral fatigue, where other factors separate to group III and IV afferents may have contributed to the down-regulation of descending central drive.

The use of stimulation (e.g. nerve, muscle, transcranial magnetic stimulation) to explore the potential link between  $\text{NaHCO}_3$  and central and peripheral mechanisms associated with skeletal muscle fatigue may provide further insight toward the ergogenic potential of this supplement. Moreover, our findings that alkalosis may affect central mechanisms associated with early rate of torque development warrants further investigation, as the ability to rapidly recruit muscle may be influenced by the

contractile properties of the fibre (e.g. type I or II) (Fig. 2) [118]. As evidence suggests that acute  $\text{NaHCO}_3$  ingestion may have a preferential effect on type II muscle fibres [83], further study comparing different combinations of exercise task demands and muscle groups may also clarify the effect of  $\text{NaHCO}_3$  on rates of torque development and muscle recruitment in humans.

## Performance Applications

### Competition and Training Recommendations

Appropriately identifying whether  $\text{NaHCO}_3$  ingestion may influence the underlying mechanisms associated with whole-body metabolism, muscle physiology and/or motor pathways within the context of a particular athletic endeavour or training stimulus is an important initial step when considering the use of this supplement. For example, in the context of athletic performance, delayed onset of  $\text{H}^+$  accumulation may be relevant to certain time frames within an event that demand rapid transitions between steady-state and higher intensity efforts (e.g. beginning a 'long-finish' earlier in a 1500-m race). In this scenario,  $\text{NaHCO}_3$  supplementation may be appropriate based on the evidence indicating an attenuation of  $\text{H}^+$  accumulation during exercise intensities transitioning between aerobic and anaerobic pathways [60, 65, 89]. Another example, and specifically related to the evidence surrounding RFD and fast-twitch fibres [83, 97], would be to improve the propulsive force at the start of an explosive movement (e.g. the initiation of a pedal stroke). Considerations for the potential benefit of  $\text{NaHCO}_3$  in these unique situations would require extensive consultation within an athlete's support network, as well as a systematic integration of the supplement to accurately determine any efficacious response. Although often difficult to rigorously assess in the elite environment, any marginal improvement in performance gained by using  $\text{NaHCO}_3$  for these specific purposes in competition may provide alternative uses for this supplement that extend beyond current recommendations [1, 5].

Although there have been few studies investigating the efficacy of implementing a chronic  $\text{NaHCO}_3$  supplementation strategy on training outcomes [89, 119], given the recent evidence in animal models exhibiting enhanced oxidative/mitochondrial adaptations in slow-twitch fibres after chronic supplementation [90, 91], further work in this area may establish whether incorporating  $\text{NaHCO}_3$  into an aerobic training paradigm is warranted. Furthermore, with research also supporting a potential ergogenic effect on RFD [97], particularly in fast-twitch fibres [83], applying a loading strategy during specific training and adaptation blocks (e.g. explosive power) may also have merit. In practice, however, incorporating a training strategy inclusive of a regular  $0.3 \text{ g}\cdot\text{kg}^{-1}$

NaHCO<sub>3</sub> load would require a systematic administration and monitored approach. Although we are unaware of any documented long-term adverse effects of chronic loading in either sporting [89, 120, 121] or clinical contexts [122–124], given the relatively high sodium content (e.g. a 70-kg athlete would consume ~6 g), continually monitoring both blood acid-base and strong ions (e.g. Na<sup>+</sup>) would allow any irregularities from the ingestion regimen to be detected and ultimately corrected for the safety of the athlete(s). Additionally, and similar to other contemporary debates such as training in a low glycogen state [125], without further study potentially elucidating any maladaptation from consecutive training blocks after NaHCO<sub>3</sub> ingestion, it would be prudent to consider intermittently incorporating this strategy and only when certain training outcomes are desired (e.g. maximising the number of repeated efforts at top speed or high rates of force output).

#### **Loading Recommendations**

Equally important to the performance and training strategies, however, may be the ingestion protocol used to systematically introduce NaHCO<sub>3</sub>. Research is only now highlighting what practitioners, coaches and athletes have been aware of for some time, and that is the inherent variability in individual responsiveness to this supplement [126–130]. In part, the variability in many papers may eventuate from the wide range of exercise tasks and methodological applications, as well as heterogeneity between participant phenotypes. However, it may also result from the widespread assumption that applying mean data on time-to-peak buffering response rates after NaHCO<sub>3</sub> ingestion prior to commencing exercise is not influencing the outcome of many performance tasks [128, 129]. Amongst others [131, 132], we have profiled the ingestion time course of various doses of NaHCO<sub>3</sub> at rest in order to determine peak response rates (Fig. 1) [3]. Collectively, and variation in ingestion protocols notwithstanding (e.g. capsules, bolus loads or dispersed over time), these studies illustrate around 40 min of variation between subjects' peak in blood buffering capacity [3, 131, 132]. Indeed, the large degree of inter-individual variability profiled in Fig. 1 has been recently observed again but under greater temporal resolution and in a larger study cohort [129]. Although we have shown that this variation may not be important for recreationally trained individuals [133], we are unaware of any studies that have documented the variability in elite athlete populations.

Finally, the origin of the 0.2 to 0.3 g·kg<sup>-1</sup> doses in the sport performance field is somewhat ambiguous. Although oral ingestion of between 25 to 40 g of NaHCO<sub>3</sub> has been documented as early as the 1920s and early 1930s in the clinical literature [124, 134, 135], this range

is believed to have come from the work of Poulus and colleagues, who in 1974 published the first exercise performance paper using a physiochemical rationale for NaHCO<sub>3</sub> dose selection [74]. Citing earlier clinical work [122, 136], these authors determined the amount of NaHCO<sub>3</sub> to be administered intravenously using the following formula:

$$\text{NaHCO}_3 = \text{subject body weight (kg)} \times 0.3 \\ \times \text{base deficit}$$

where base deficit (BD) represented the excess of fixed acid in the blood measured after an incremental cycling test on a previous visit to the lab and compared to a standard (pH of 7.4 and a PCO<sub>2</sub> of 40 mmHg) [74]. The 0.3 represented a mean weighting factor derived from the equilibrating time course of NaHCO<sub>3</sub> throughout blood and the extravascular spaces [122, 123]. Of note, no subsequent studies have included BD in the equation for determining dose. This, coupled with the variation in individual weighting factors reported in the original study (0.25 to 0.38) [122], warrants further work to determine whether or not individualising dose concentrations is necessary, and whether issues related to HCO<sub>3</sub><sup>-</sup> and [H<sup>+</sup>] rates of increase and clearance are impacting upon the efficacy of NaHCO<sub>3</sub> as a supplement [130].

The dosing papers most frequently cited in the sport performance literature date back to the late 1980s and early 1990s. These studies looked at a range of NaHCO<sub>3</sub> doses (0.1 to 0.5 g·kg<sup>-1</sup>) and the relationship between performance and gastrointestinal (GI) disturbance, respectively [137, 138]. Collectively, these studies showed that performance could be improved when the acute dose ingested ranged between 0.2 and 0.3 g·kg<sup>-1</sup>, but anything greater was not beneficial and only induced severe side effects. For the past 30 years, this dose range has been widely accepted as best practice. Yet, the small subject numbers ( $n < 9$ ) coupled with their non-elite training status (e.g. 'participating in athletic events') of the original research should warrant further inquiry [138]. This, of course, also assumes that 0.2 or 0.3 g·kg<sup>-1</sup> are the optimal physiologic doses for maximising blood buffering capacity [130].

In light of the practical issues identified in this section, we recommend monitoring at an individual level the time-to-peak rise (and decay [130]) in [HCO<sub>3</sub><sup>-</sup>] at doses between 0.2 and 0.3 g·kg<sup>-1</sup> and to tailor the commencement of exercise accordingly [128]. An individualised loading strategy will also require systematically incorporating dose distribution and other planned nutritional interventions, documenting subjective feedback from the athlete (e.g. gastrointestinal distress [139]) and monitoring any physiological changes (e.g.

acute weight gain from plasma volume expansion [21]) observed after ingestion.

## Conclusions

We have attempted to provide a comprehensive overview of the mechanistic links between skeletal muscle fatigue, metabolic acidosis and NaHCO<sub>3</sub> supplementation. The synergistic associations presented within the 'Whole-body Metabolism', 'Muscle Physiology' and 'Motor Pathways' sections provide evidence that an integrative perspective is required to truly understand and optimise the use and application of this supplement. By consolidating the mechanistic evidence into one text, we aimed to provide new insights and possibly direct future research in the field of NaHCO<sub>3</sub> supplementation. Finally, we hope that this review provides further evidence base to expand and refine the use of this supplement to improve athletic performance beyond traditional methods of application.

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## Authors' contributions

JS is responsible for the conceptual framework and design, literature review, manuscript preparation, critical revision and approval of the review. PM, DB and GS contributed to the manuscript preparation, critical revision and approval of the review. SG contributed to the literature review, manuscript preparation, critical revision and approval of the review.

## Competing interests

Jason Siegler, Paul Marshall, David Bishop, Greg Shaw and Simon Green declare that they have no conflict of interest.

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## References

- Carr AJ, Hopkins WG, Gore CJ. Effects of acute alkalosis and acidosis on performance: a meta-analysis. *Sports Med.* 2011;41:801–14.
- Pearl DJ, Siegler JC, Vince RV. Practical recommendations for coaches and athletes: a meta-analysis of sodium bicarbonate use for athletic performance. *J Strength Cond Res.* 2012;26:1975–83.
- Siegler JC, Midgley AW, Polman RCJ, Lever R. Effects of various sodium bicarbonate loading protocols on the time-dependent extracellular buffering profile. *J Strength Cond Res.* 2010;24:2551–7.
- Mcnaughton LR, Siegler J, Midgley A. Ergogenic effects of sodium bicarbonate. *Curr Sports Med Rep.* 2008;7:230–6.
- Mcnaughton LR, Gough L, Deb S, Bentley D, Sparks SA. Recent developments in the use of sodium bicarbonate as an ergogenic aid. *Curr Sports Med Rep.* 2016;15:233–44.
- Spriet LL, Matsos CG, Peters SJ, Heigenhauser GJ, Jones NL. Effects of acidosis on rat muscle metabolism and performance during heavy exercise. *Am J Physiol.* 1985;248:C337–47.
- Spriet L, Lindinger M, McKelvie R, Heigenhauser G, Jones N. Muscle glycogenolysis and H<sup>+</sup> concentration during maximal intermittent cycling. *J Appl Physiol.* 1989;66:8–13.
- Knuth ST, Dave H, Peters JR, Fitts RH. Low cell pH depresses peak power in rat skeletal muscle fibres at both 30 degrees C and 15 degrees C: implications for muscle fatigue. *J Physiol.* 2006;575:887–99.
- Debold EP, Beck SE, Warshaw DM. Effect of low pH on single skeletal muscle myosin mechanics and kinetics. *Am J Physiol Cell Physiol.* 2008;295:C173–9.
- Westerblad H, Allen DG. Changes of intracellular pH due to repetitive stimulation of single fibres from mouse skeletal muscle. *J Physiol.* 1992;449:49–71.
- Overgaard K, Højfeldt GW, Nielsen OB. Effects of acidification and increased extracellular potassium on dynamic muscle contractions in isolated rat muscles. *J Physiol.* 2010;588:5065–76.
- Kent-Braun JA, Fitts RH, Christie A. Skeletal muscle fatigue. *Compr Physiol.* 2011;2:997–1044.
- Cairns SP, Lindinger ML. Do multiple ionic interactions contribute to skeletal muscle fatigue? *J Physiol.* 2008;586:4039–54.
- Debold EP. Recent insights into the molecular basis of muscular fatigue. *Med Sci Sports Exerc.* 2012;44:1440–52.
- Bishop D, Claudius B. Effects of induced metabolic alkalosis on prolonged intermittent-sprint performance. *Med Sci Sports Exerc.* 2005;37:759–67.
- Cameron SL, McIay-Cooke RT, Brown RC, Gray AR, Fairbairn KA. Increased blood pH but not performance with sodium bicarbonate supplementation in elite rugby union players. *Int J Sport Nutr Exerc Metab.* 2010;20:307–21.
- Price M, Moss P, Rance S. Effects of sodium bicarbonate ingestion on prolonged intermittent exercise. *Med Sci Sports Exerc.* 2003;35:1303–8.
- Artoli GG, Gualano B, Coelho DF, Benatti FB, Gailey AW, Lancha AH. Does sodium-bicarbonate ingestion improve simulated judo performance? *Int J Sport Nutr Exerc Metab.* 2007;17:206–17.
- Siegler JC, Hirscher K. Sodium bicarbonate ingestion and boxing performance. *J Strength Cond Res.* 2010;24:103–8.
- Wu C-L, Shih M-C, Yang C-C, Huang M-H, Chang C-K. Sodium bicarbonate supplementation prevents skilled tennis performance decline after a simulated match. *J Int Soc Sports Nutr.* 2010;7:33.
- Kupcis PD, Slater GJ, Pruscino CL, Kemp JG. Influence of sodium bicarbonate on performance and hydration in lightweight rowing. *Int J Sports Physiol Perform.* 2012;7:11–8.
- Mitchell TH, Abraham G, Wing S, Magder SA, Cosio MG, Deschamps A, et al. Intravenous bicarbonate and sodium chloride both prolong endurance during intense cycle ergometer exercise. *Am J Med Sci.* 1990;300:88–97.
- Gonzalez NC, Zamagni M, Clancy RL. Effect of alkalosis on maximum oxygen uptake in rats acclimated to simulated altitude. *J Appl Physiol.* 1991;71:1050–6.
- Kozak-Collins K, Burke ER, Schoene RB. Sodium bicarbonate ingestion does not improve performance in women cyclists. *Med Sci Sports Exerc.* 1994;26:1510–5.
- Flinn S, Herbert K, Graham K, Siegler JC. Differential effect of metabolic alkalosis and hypoxia on high-intensity cycling performance. *J Strength Cond Res.* 2014;28:2852–8.
- Dennig H, Talbott JH, Edwards HT, Dill DB. Effect of acidosis and alkalosis upon capacity for work. *J Clin Invest.* 1931;9:601–13.
- Needham DM. *Machina Carnis*. Cambridge: Cambridge University Press; 2009.
- Asmussen E, Döbeln WV, Nielsen M. Blood lactate and oxygen debt after exhaustive work at different oxygen tensions. *Acta Physiol Scand.* 1948;15:57–62.
- Johnson WR, Black DH. Comparison of effects of certain blood alkalizers and glucose upon competitive endurance performance. *J Appl Physiol.* 1953;5:577–8.
- Margaria R, Aghemo P, Sassi G. Effect of alkalosis on performance and lactate formation in supramaximal exercise. *Int Z Angew Physiol.* 1971;29:215–23.
- Simmons RW, Hardt AB. The effect of alkali ingestion on the performance of trained swimmers. *J Sports Med Phys Fitness.* 1973;13:159–63.
- Jones NL, Sutton JR, Taylor R, Toews CJ. Effect of pH on cardiorespiratory and metabolic responses to exercise. *J Appl Physiol Respir Environ Exerc Physiol.* 1977;43:959–64.
- Sutton JR, Jones NL, Toews CJ. Effect of pH on muscle glycolysis during exercise. *Clin Sci.* 1981;61:331–8.
- McCartney N, Heigenhauser GJ, Jones NL. Effects of pH on maximal power output and fatigue during short-term dynamic exercise. *J Appl Physiol Respir Environ Exerc Physiol.* 1983;55:225–9.

35. Kowalchuk JM, Heigenhauser GJ, Jones NL. Effect of pH on metabolic and cardiorespiratory responses during progressive exercise. *J Appl Physiol Respir Environ Exerc Physiol.* 1984;57:1558–63.
36. Wilkes D, Gledhill N, Smyth R. Effect of acute induced metabolic alkalosis on 800-m racing time. *Med Sci Sports Exerc.* 1983;15:277–80.
37. Gao JP, Costill DL, Horswill CA, Park SH. Sodium bicarbonate ingestion improves performance in interval swimming. *Eur J Appl Physiol Occup Physiol.* 1988;58:171–4.
38. McKenzie DC, Coutts KD, Stirling DR, Hoeben HH, Kuzara G. Maximal work production following two levels of artificially induced metabolic alkalosis. *J Sports Sci.* 1986;4:35–8.
39. Goldfinch J, Mc Naughton L, Davies P. Induced metabolic alkalosis and its effects on 400-m racing time. *Eur J Appl Physiol Occup Physiol.* 1988;57:45–8.
40. Costill DL, Verstappen F, Kuipers H, Janssen E, Fink W. Acid-base balance during repeated bouts of exercise: influence of  $\text{HCO}_3^-$ . *Int J Sports Med.* 1984;5:228–31.
41. Linderman JK, Gosselink KL. The effects of sodium bicarbonate ingestion on exercise performance. *Sports Med.* 1994;18:75–80.
42. Matson LG, Tran ZV. Effects of sodium bicarbonate ingestion on anaerobic performance: a meta-analytic review. *Int J Sport Nutr.* 1993;3:2–28.
43. George KP, Maclaren DPM. The effect of induced alkalosis and acidosis on endurance running at an intensity corresponding to 4 mM blood lactate. *Ergonomics.* 1988;31(11):1639–45.
44. Brien DM, McKenzie DC. The effect of induced alkalosis and acidosis on plasma lactate and work output in elite oarsmen. *Eur J Appl Physiol Occup Physiol.* 1989;58:797–802.
45. Lavender G, Bird SR. Effect of sodium bicarbonate ingestion upon repeated sprints. *Br J Sports Med.* 1989;23:41–5.
46. Duncan MJ, Weldon A, Price MJ. The effect of sodium bicarbonate ingestion on back squat and bench press exercise to failure. *J Strength Cond Res.* 2014;28:1358–66.
47. Egger F, Meyer T, Such U, Hecksteden A. Effects of sodium bicarbonate on high-intensity endurance performance in cyclists: a double-blind, randomized cross-over trial. *PLoS ONE.* 2014;9:e114729–15.
48. Krstrup P, Ermidis G, Mohr M. Sodium bicarbonate intake improves high-intensity intermittent exercise performance in trained young men. *J Int Soc Sports Nutr.* 2015;1–7.
49. Hobson RM, Harris RC, Martin D, Smith P, Macklin B, Elliott-Sale KJ, et al. Effect of sodium bicarbonate supplementation on 2000-m rowing performance. *Int J Sports Physiol Perform.* 2014;9:139–44.
50. Berger NJA, Mcnaughton LR, Keatley S, Wilkerson DP, Jones AM. Sodium bicarbonate ingestion alters the slow but not the fast phase of  $\text{VO}_2$  kinetics. *Med Sci Sports Exerc.* 2006;38:1909–17.
51. Zoladz JA, Szkutnik Z, Krzysztof D, Majerczak J, Korzeniewski B. Preexercise metabolic alkalosis induced via bicarbonate ingestion accelerates  $\text{VO}_2$  kinetics at the onset of a high-power-output exercise in humans. *J Appl Physiol.* 2004;98:895–904.
52. Kolkhorst FW, Rezende RS, Levy SS, Buono MJ. Effects of sodium bicarbonate on  $\text{VO}_2$  kinetics during heavy exercise. *Med Sci Sports Exerc.* 2004;36:1895–9.
53. Hermansen L, Osnes JB. Blood and muscle pH after maximal exercise in man. *J Appl Physiol.* 1972;32:304–8.
54. Mainwood GW, Worsley-Brown P. The effects of extracellular pH and buffer concentration on the efflux of lactate from frog sartorius muscle. *J Physiol.* 1975; 250:1–22.
55. Hirche HJ, Hombach V, Langohr HD, Wacker U, Busse J. Lactic acid permeation rate in working gastrocnemii of dogs during metabolic alkalosis and acidosis. *Pflügers Arch Eur J Physiol.* 1975;356:209–22.
56. Spriet LL, Lindinger MI, Heigenhauser GJ, Jones NL. Effects of alkalosis on skeletal muscle metabolism and performance during exercise. *Am J Physiol.* 1986;251:R833–9.
57. Hollidge-Horvat MG, Parolin ML, Wong D, Jones NL, Heigenhauser GJ. Effect of induced metabolic alkalosis on human skeletal muscle metabolism during exercise. *Am J Physiol Endocrinol Metab.* 2000;278:E316–29.
58. Granier PL, Dubouchaud H, Mercier BM, Mercier JG, Ahmaidi S, Préfaut CG. Effect of  $\text{NaHCO}_3$  on lactate kinetics in forearm muscles during leg exercise in man. *Med Sci Sports Exerc.* 1996;28:692–7.
59. Bishop D, Edge J, Thomas C, Mercier J. High-intensity exercise acutely decreases the membrane content of MCT1 and MCT4 and buffer capacity in human skeletal muscle. *J Appl Physiol.* 2006;102:616–21.
60. Raymer GH, Marsh GD, Kowalchuk JM, Thompson RT. Metabolic effects of induced alkalosis during progressive forearm exercise to fatigue. *J Appl Physiol.* 2004;96:2050–6.
61. Forbes SC, Raymer GH, Kowalchuk JM, Marsh GD.  $\text{NaHCO}_3$ -induced alkalosis reduces the phosphocreatine slow component during heavy-intensity forearm exercise. *J Appl Physiol.* 2005;99:1668–75.
62. Churchward-Venne TA, Kowalchuk JM, Marsh GD. Effects of ammonium chloride ingestion on phosphocreatine metabolism during moderate- and heavy-intensity plantar-flexion exercise. *Eur J Appl Physiol.* 2009;108:1189–200.
63. Spriet LL, Soderlund K, Bergstrom M, Hultman E. Skeletal muscle glycogenolysis, glycolysis, and pH during electrical stimulation in men. *J Appl Physiol.* 1987;62:616–21.
64. Spriet LL, Lindinger MI, Mckelvie RS, Heigenhauser GJ, Jones NL. Muscle glycogenolysis and  $\text{H}^+$  concentration during maximal intermittent cycling. *J Appl Physiol.* 1989;66:8–13.
65. Sostaric SM, Skinner TSL, Brown MJ, Sangkabutra T, Medved I, Medley T, et al. Alkalosis increases muscle  $\text{K}^+$  release, but lowers plasma  $[\text{K}^+]$  and delays fatigue during dynamic forearm exercise. *J Physiol.* 2005;570:185–205.
66. Street D, Nielsen JJ, Bangsbo J, Juel C. Metabolic alkalosis reduces exercise-induced acidosis and potassium accumulation in human skeletal muscle interstitium. *J Physiol.* 2005;566:481–9.
67. Bouissou P, Defer G, Guezennec CY, Estrade PY, Serrurier B. Metabolic and blood catecholamine responses to exercise during alkalosis. *Med Sci Sports Exerc.* 1988;20:228–32.
68. Bird SR, Wiles J, Robbins J. The effect of sodium bicarbonate ingestion on 1500-m racing time. *J Sports Sci.* 1995;13:399–403.
69. van Montfoort MCE, van Dieren L, Hopkins WG, Shearman JP. Effects of ingestion of bicarbonate, citrate, lactate, and chloride on sprint running. *Med Sci Sports Exerc.* 2004;36:1239–43.
70. Lindh A, Peyrebrune M, Ingham S, Bailey D, Folland J. Sodium bicarbonate improves swimming performance. *Int J Sports Med.* 2008;29:519–23.
71. Broch-Lips M, Overgaard K, Praetorius HA, Nielsen OB. Effects of extracellular  $\text{HCO}_3^-$  on fatigue,  $\text{pH}_i$ , and  $\text{K}^+$  efflux in rat skeletal muscles. *J Appl Physiol.* 2007;103:494–503.
72. Sejersted OM, Sjøgaard G. Dynamics and consequences of potassium shifts in skeletal muscle and heart during exercise. *Physiol Rev.* 2000;80:1411–81.
73. Houtman CJ, Heerschap A, Zwarts MJ, Stegeman DF. pH heterogeneity in tibial anterior muscle during isometric activity studied by  $^{31}\text{P}$ -NMR spectroscopy. *J Appl Physiol.* 2001;91:191–200.
74. Poulos AJ, Docter HJ, Westra HG. Acid-base balance and subjective feelings of fatigue during physical exercise. *Eur J Appl Physiol Occup Physiol.* 1974;33:207–13.
75. Housh TJ, de Vries HA, Johnson GO, Evans SA, McDowell S. The effect of ammonium chloride and sodium bicarbonate ingestion on the physical working capacity at the fatigue threshold. *Eur J Appl Physiol Occup Physiol.* 1991;62:189–92.
76. Fabiato A, Fabiato F. Effects of pH on the myofilaments and the sarcoplasmic reticulum of skinned cells from cardiac and skeletal muscles. *J Physiol.* 1978;276:233–55.
77. Edman KA, Mattiazzi AR. Effects of fatigue and altered pH on isometric force and velocity of shortening at zero load in frog muscle fibres. *J Muscle Res Cell Motil.* 1981;2:321–34.
78. Robertson SP, Glenn W, Kerrick L. The effects of pH on  $\text{Ca}^{2+}$ -activated force in frog skeletal muscle fibres. *Pflügers Arch Eur J Physiol.* 1979;380:41–5.
79. Metzger JM, Moss RL. Greater hydrogen ion-induced depression of tension and velocity in skinned single fibres of rat fast than slow muscles. *J Physiol.* 1987;393:727–42.
80. Baldwin KM, Klinkerkuff GH, Terjung RL, Molé PA, Holloszy JO. Respiratory capacity of white, red, and intermediate muscle: adaptive response to exercise. *Am J Physiol.* 1972;222:373–8.
81. Ariano MA, Edgerton VR, Armstrong RB. Hindlimb muscle fiber populations of five mammals. *J Histochem Cytochem.* 1973;21:51–5.
82. Lindinger MI, Heigenhauser GJ, Spriet LL. Effects of alkalosis on muscle ions at rest and with intense exercise. *Can J Physiol Pharmacol.* 1990;68:820–9.
83. Higgins MF, Tallis J, Price MJ, James RS. The effects of elevated levels of sodium bicarbonate ( $\text{NaHCO}_3$ ) on the acute power output and time to fatigue of maximally stimulated mouse soleus and EDL muscles. *Eur J Appl Physiol.* 2012;113:1331–41.
84. Inbar O, Rotstein A, Jacobs I, Kaiser P, Dlin R, Dotan R. The effects of alkaline treatment on short-term maximal exercise. *J Sports Sci.* 1983;1:95–104.
85. Zhang SJ, Bruton JD, Katz A, Westerblad H. Limited oxygen diffusion accelerates fatigue development in mouse skeletal muscle. *J Physiol.* 2006;572:551–9.
86. Benadé AJ, Heisler N. Comparison of efflux rates of hydrogen and lactate ions from isolated muscles in vitro. *Respir Physiol.* 1978;32:369–80.

87. Webster MJ, Webster MN, Crawford RE, Gladden LB. Effect of sodium bicarbonate ingestion on exhaustive resistance exercise performance. *Med Sci Sports Exerc.* 1993;25:960–5.
88. Portington KJ, Pascoe DD, Webster MJ, Anderson LH, Rutland RR, Gladden LB. Effect of induced alkalosis on exhaustive leg press performance. *Med Sci Sports Exerc.* 1998;30:523–8.
89. Edge J, Bishop D, Goodman C. Effects of chronic NaHCO<sub>3</sub> ingestion during interval training on changes to muscle buffer capacity, metabolism, and short-term endurance performance. *J Appl Physiol.* 2006;101:918–25.
90. Thomas C, Bishop D, Moore-Morris T, Mercier J. Effects of high-intensity training on MCT1, MCT4, and NBC expressions in rat skeletal muscles: influence of chronic metabolic alkalosis. *Am J Physiol Endocrinol Metab.* 2007;293:E916–22.
91. Bishop DJ, Thomas C, Moore-Morris T, Tonkonogi M, Sahlin K, Mercier J. Sodium bicarbonate ingestion prior to training improves mitochondrial adaptations in rats. *Am J Physiol Endocrinol Metab.* 2010;299:E225–33.
92. Bouissou P, Estrade PY, Goubel F, Guezennec CY, Serrurier B. Surface EMG power spectrum and intramuscular pH in human vastus lateralis muscle during dynamic exercise. *J Appl Physiol.* 1989;67:1245–9.
93. Hunter AM, De Vito G, Bolger C, Mullany H, Galloway SDR. The effect of induced alkalosis and submaximal cycling on neuromuscular response during sustained isometric contraction. *J Sports Sci.* 2009;27:1261–9.
94. Matsuura R, Arimitsu T, Kimura T, Yunoki T, Yano T. Effect of oral administration of sodium bicarbonate on surface EMG activity during repeated cycling sprints. *Eur J Appl Physiol.* 2007;101:409–17.
95. Yamanaka R, Yunoki T, Arimitsu T, Lian C-S, Yano T. Effects of sodium bicarbonate ingestion on EMG, effort sense and ventilatory response during intense exercise and subsequent active recovery. *Eur J Appl Physiol.* 2010;111:851–8.
96. Lowery M, Nolan P, O'Malley M. Electromyogram median frequency, spectral compression and muscle fibre conduction velocity during sustained sub-maximal contraction of the brachioradialis muscle. *J Electromyogr Kinesiol.* 2002;12:111–8.
97. Siegler JC, Marshall PWM, Raftoy S, Brooks C, Dowsell B, Romero R, et al. The differential effect of metabolic alkalosis on maximum force and rate of force development during repeated, high-intensity cycling. *J Appl Physiol.* 2013;115:1634–40.
98. Aagaard P, Simonsen EB, Andersen JL, Magnusson P, Dyhre-Poulsen P. Increased rate of force development and neural drive of human skeletal muscle following resistance training. *J Appl Physiol.* 2002;93:1318–26.
99. Gandevia SC. Spinal and supraspinal factors in human muscle fatigue. *Physiol Rev.* 2001;81:1725–89.
100. Amann M. Central and peripheral fatigue. *Med Sci Sports Exerc.* 2011;43:2039–45.
101. Beekley MD. Carbon dioxide alters the Hoffmann reflex independent of hydrogen ions. *Int J Neurosci.* 2014;124:518–23.
102. Kniffki KD, Schomburg ED, Steffens H. Synaptic responses of lumbar alpha-motoneurons to chemical algescic stimulation of skeletal muscle in spinal cats. *Brain Res.* 1979;160:549–52.
103. Rotto DM, Kaufman MP. Effect of metabolic products of muscular contraction on discharge of group III and IV afferents. *J Appl Physiol.* 1988;64:2306–13.
104. Taylor JL, Butler JE, Gandevia SC. Changes in muscle afferents, motoneurons and motor drive during muscle fatigue. *Eur J Appl Physiol.* 2000;83:106–15.
105. Rossi A, Mazzocchio R, Decchi B. Effect of chemically activated fine muscle afferents on spinal recurrent inhibition in humans. *Clin Neurophysiol.* 2003;114:279–87.
106. Fischer M, Schäfer SS. Effects of changes in pH on the afferent impulse activity of isolated cat muscle spindles. *Brain Res.* 2005;1043:163–78.
107. Søgaard K, Gandevia SC, Todd G, Petersen NT, Taylor JL. The effect of sustained low-intensity contractions on supraspinal fatigue in human elbow flexor muscles. *J Physiol.* 2006;573:511–23.
108. Adam A, De Luca CJ. Firing rates of motor units in human vastus lateralis muscle during fatiguing isometric contractions. *J Appl Physiol.* 2005;99:268–80.
109. Dideriksen JL, Enoka RM, Farina D. Neuromuscular adjustments that constrain submaximal EMG amplitude at task failure of sustained isometric contractions. *J Appl Physiol.* 2011;111:485–94.
110. Light AR, Hughen RW, Zhang J, Rainier J, Liu Z, Lee J. Dorsal root ganglion neurons innervating skeletal muscle respond to physiological combinations of protons, ATP, and lactate mediated by ASIC, P2X, and TRPV1. *J Neurophysiol.* 2008;100:1184–201.
111. Jankowski MP, Rau KK, Ekmann KM, Anderson CE, Koerber HR. Comprehensive phenotyping of group III and IV muscle afferents in mouse. *J Neurophysiol.* 2013;109:2374–81.
112. Pollak KA, Swenson JD, Vanhaisma TA, Hughen RW, Jo D, White AT, et al. Exogenously applied muscle metabolites synergistically evoke sensations of muscle fatigue and pain in human subjects. *Ex Physiol.* 2014;99:368–80.
113. Amann M, Venturelli M, Ives SJ, McDaniel J, Layec G, Rossman MJ, et al. Peripheral fatigue limits endurance exercise via a sensory feedback-mediated reduction in spinal motoneuronal output. *J Appl Physiol.* 2013;115:355–64.
114. Taylor JL, Petersen N, Butler JE, Gandevia SC. Schaemia after exercise does not reduce responses of human motoneurons to cortical or corticospinal tract stimulation. *J Physiol.* 2000;525:793–801.
115. Martin PG. Fatigue-sensitive afferents inhibit extensor but not flexor motoneurons in humans. *J Neurosci.* 2006;26:4796–802.
116. Rossman MJ, Garten RS, Venturelli M, Amann M, Richardson RS. The role of active muscle mass in determining the magnitude of peripheral fatigue during dynamic exercise. *Am J Physiol Regul Integr Comp Physiol.* 2014;306:R934–40.
117. Siegler JC, Marshall P. The effect of metabolic alkalosis on central and peripheral mechanisms associated with exercise-induced muscle fatigue in humans. *Ex Physiol.* 2015;100:519–30.
118. Van Cutsem M, Duchateau J, Hainaut K. Changes in single motor unit behaviour contribute to the increase in contraction speed after dynamic training in humans. *J Physiol.* 1998;513(Pt1):295–305.
119. Driller MW, Gregory JR, Williams AD, Fell JW. The effects of chronic sodium bicarbonate ingestion and interval training in highly trained rowers. *Int J Sport Nutr Exerc Metab.* 2013;23:40–7.
120. McNaughton L, Backx K, Palmer G, Strange N. Effects of chronic bicarbonate ingestion on the performance of high-intensity work. *Eur J Appl Physiol Occup Physiol.* 1999;80:333–6.
121. Douroudos II, Fatouros IG, Gourgoulis V, Jamurtas AZ, Tsitsios T, Hatzinikolaou A, et al. Dose-related effects of prolonged NaHCO<sub>3</sub> ingestion during high-intensity exercise. *Med Sci Sports Exerc.* 2006;38:1746–53.
122. Mellemegaard K, Astrup P. The quantitative determination of surplus amounts of acid or base in the human body. *Scand J Clin Lab Invest.* 1960;12:187–99.
123. Singer RB, Clark JK, Barker ES, Crosley AP, Elkinton JR. The acute effects in man of rapid intravenous infusion of hypertonic sodium bicarbonate solution. I. Changes in acid-base balance and distribution of the excess buffer base. *Medicine (Baltimore).* 1955;34:51–95.
124. Shock NW, Hastings AB. Studies of the acid-base balance of the blood. *J Biol Chem.* 1935;112:239–62.
125. Hawley JA, Morton JP. Ramping up the signal: promoting endurance training adaptation in skeletal muscle by nutritional manipulation. *Clin Exp Pharmacol Physiol.* 2014;41:608–13.
126. de Araujo Dias FG, da Eira S, de Salles P, Sale C, Artioli GG, Gualano B, et al. (In)consistencies in responses to sodium bicarbonate supplementation: a randomised, repeated measures, counterbalanced and double-blind study. *PLoS One.* 2015;10(11):e0143086.
127. Saunders B, Sale C, Harris RC, Sunderland C. Sodium bicarbonate and high-intensity-cycling capacity: variability in responses. *Int J Sports Physiol Perform.* 2014;9:627–32.
128. Miller P, Robinson AL, Sparks SA, Bridge CA, Bentley DJ, Mcnaughton LR. The effects of novel ingestion of sodium bicarbonate on repeated sprint ability. *J Strength Cond Res.* 2016;30:561–8.
129. Stannard RL, Stellingwerff T, Artioli GG, Saunders B, Cooper S, Sale C. Dose-response of sodium bicarbonate ingestion highlights individuality in time course of blood analyte responses. *Int J Sport Nutr Exerc Metab.* 2016;1–20.
130. Green S, Siegler JC. Empirical modeling of metabolic alkalosis induced by sodium bicarbonate ingestion. *Appl Physiol Nutr Metab.* 2016. <http://dx.doi.org/10.1123/ijnsnem.2015-0286>.
131. Renfree A. The time course for changes in plasma [H<sup>+</sup>] after sodium bicarbonate ingestion. *Int J Sports Physiol Perform.* 2007;2:323–6.
132. Price MJ, Singh M. Time course of blood bicarbonate and pH three hours after sodium bicarbonate ingestion. *Int J Sports Physiol Perform.* 2008;3:240–2.
133. Siegler JC, Marshall PWM, Bray J, Towson C. Sodium bicarbonate supplementation and ingestion timing: does it matter? *J Strength Cond Res.* 2012;26:1953–8.
134. Koehler AE. The effect of acid and base ingestion upon the acid-base balance. *J Biol Chem.* 1927;72:99–122.

135. Palmer WW, Salvesen H, Jackson HJ. Relationship between the plasma bicarbonate and urinary acidity following the administration of sodium bicarbonate. *J Biol Chem.* 1920;45:101–11.
136. Astrup P, Jørgensen K, Andersen OS, Engel K. The acid-base metabolism. A new approach. *Lancet.* 1960;1:1035–9.
137. Horswill CA, Costill DL, Fink WJ, Flynn MG, Kirwan JP, Mitchell JB, et al. Influence of sodium bicarbonate on sprint performance: relationship to dosage. *Med Sci Sports Exerc.* 1988;20:566–9.
138. Mcnaughton LR. Bicarbonate ingestion: effects of dosage on 60 s cycle ergometry. *J Sports Sci.* 1992;10:415–23.
139. Carr AJ, Slater GJ, Gore CJ, Dawson B, Burke LM. Effect of sodium bicarbonate on  $[\text{HCO}_3^-]$ , pH, and gastrointestinal symptoms. *Int J Sport Nutr Exerc Metab.* 2011;2011:1–6.

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